NAUTIAGENE

Product Information

NautiaZ Tissue DNA Extraction Mini Kit

(100/300 prep)

Cat. No.: NGTZ-S100/NGTZ-S300

Sample: 30 mg of fresh animal tissue

25 mg of paraffin embedded tissue

Yield: Up to 50 μg

NGCZ-S100	NautiaZ Culture Cell DNA Extraction Mini Kit (100 prep)
NGCZ-S300	NautiaZ Culture Cell DNA Extraction Mini Kit (300 prep)
NGBZ-S100	NautiaZ Blood DNA Extraction Mini Kit (100 prep)
NGBZ-S300	NautiaZ Blood DNA Extraction Mini Kit (300 prep)
NGBAZ-S100	NautiaZ Bacteria/Fungi DNA Extraction Mini Kit (100 prep)
NGBAZ-S300	NautiaZ Bacteria/Fungi DNA Extraction Mini Kit (300 prep)
NGPZ-S100	NautiaZ Plant DNA Extraction Mini Kit (100 prep)
NGPZ-S300	NautiaZ Plant DNA Extraction Mini Kit (300 prep)
NGTZ-S100	NautiaZ Tissue DNA Extraction Mini Kit (100 prep)
NGTZ-S300	NautiaZ Tissue DNA Extraction Mini Kit (300 prep)
NGST-S100	NautiaZ Stool/Soil DNA Extraction Mini Kit (100 prep)
NGST-S300	NautiaZ Stool/Soil DNA Extraction Mini Kit (300 prep)

NGCF-S100	NautiaZ Cell-Free DNA Extraction Mini Kit (100 prep)
NGPK-S100	NautiaZ Whole Blood DNA Extraction Mini Kit (100 prep)

Contents

	NGTZ-S100T	NGTZ-S100	NGTZ-S300
T1 Buffer	1.5 ml	35 ml	95 ml
T2 Buffer	0.5 ml	12 ml	35 ml
W1 Buffer	2 ml	45 ml	125 ml
W2 Buffer*	300 ul x2	15 ml	25 ml x2
Elution Buffer	1 ml	10 ml	30 ml
GZ Column	4 pcs	100 pcs	300 pcs
Collection Tube	4 pcs	100 pcs	300 pcs
User Manual	1	1	1

^{*}Add 1.2 ml x2 / 60 ml / 100 ml x2 ethanol (96-100%) to W2 Buffer prior to the initial use.

Buffer Preparation

• Add ethanol (96-100%) to the Wash Solution prior to first use

	NGTZ-S100T	NGTZ-S100	NGTZ-S300
W2 Buffer	300 ul x2	15 ml	25 ml x2
ethanol (96 ~ 100%)	1.2 ml x2	60 ml	100 ml x2

Important Notes

- 1. Buffers provided in this system contain irritants. Wear gloves and lab coat when handling these buffers.
- 2. Add ethanol (96-100 %) to W2 Buffer when first open.
- 3. Prepare dry baths or water baths before the operation.
- 4. Resolve any precipitate by warming at 37°C.

Description

The NautiaZ Tissue DNA Extraction Mini Kit is designed for rapid extraction of pure genomic DNA from fresh animal tissue or paraffin-embedded tissue. Efficiently remove cellular debris and inhibitors, this kit using column-type tube in purification process through three simple steps of binding, washing and then elution for the safe and convenient extraction of high-purity genomic DNA. The entire process can be completed in less than 1 hour without phenol/chloroform, and the final product can be used in PCR or other downstream experiments.`

Storage

Store at room temperature.





ind the sample under uid nitrogen to a fine wder.

Add 100 µl T2 Buffer Centrifuge for 3 min Transfer supernatant to new 1.5 ml tube

Add 300 µl absolute ethanol vortexing.



Centrifuge for 30 sec Discard the flow-through

Add 400 µl W1 Buffer

Add 600 µl W2 Buffer Centrifuge for 30 sec Discard the flow-through



Add 50-200 μl Elution Buffer Stand at 75°C for 3 mins

Centrifuge for 2 mins to elution DNA

Prepare



until it is completely dissolved.

Cell Lysis



Binding

STEP



Discard the flow-through

Wash

PROCEDURE







Elution

For Fresh Tissue

STEP	PROCEDURE
1 Sample prepare	Cut off 30 mg of fresh animal tissue. Grind the sample under liquid nitrogen to a fine powder with pestle and mortar.

For Paraffin embedded Tissue

STEP	PROCEDURE
1 Sample prepare	Slice up to 25 mg of paraffin-embedded tissue and transfer to a 1.5 ml tube. Add 1 ml xylene and vortex vigorously and incubate at room temperature for 10 minutes. Vortex every 2 minutes during incubation.
	Centrifuge 3 minutes at 14,000 x g. Remove the supernatant. Add 1 ml absolute ethanol and mix by inverting.
	Centrifuge 3 minutes at 14,000 x g. Remove the supernatant. Add 1 ml absolute ethanol and mix by inverting.
	Centrifuge 3 minutes at 14,000 x g. Remove the supernatant. Open the tube and incubate at 37°C for 15 minutes.

PURIFICATION PROTOCOLS

SIEP	PROCEDURE
2 Cell Lysis	Add 300 µl T1 Buffer and 20 µl Proteinase K (10mg/ml) to the sample. Incubate at 60°C for 30 mins until the sample lysate is clear. Invert the tube every 5 minutes during incubation. Pre-heat the Elution Buffer at 60°C.
Optional Step: RNase	If RNA-free genomic DNA is required, perform this optional step. Add 5 µl of RNase (10 mg/ ml) to sample lysate and mix by vortexing. Incubate at room temperature for 5 mins.
3 Protein Removal	Add 100 µl T2 Buffer to the sample and shake vigorously. Centrifuge 3 mins at 14,000 x g. Transfer the supernatant to a new 1.5 ml tube. Add 300 µl absolute ethanol and shake vigorously.
4 DNA Binding	Place a GZ column in Collection Tube. Transfer the sample mixture to GZ column and centrifuge 30 seconds at 14,000 x g. Discard the flow-through and place GZ Column back in the Collection tube.
5-1 Wash	Add 400 μ l W1 Buffer to GZ Column. Centrifuge at 14,000 x g for 30 seconds. Discard the flow-through and place GZ Column back in the Collection tube.

5-2 Wash	Add 600 μ I W2 Buffer (ethanol added) to GZ Column. Centrifuge at 14,000 x g for 30 seconds. Discard the flow-through and place GZ Column back in the Collection tube.
6 Dry	Centrifuge at $14,000 \times g$ for 2 minutes to dry the column.
7 Elution	Place GZ Column to a clean 1.5 ml microcentrifuge tube (not provided). Add 50-200 μl of preheated Elution Buffer into the center of the column matrix. Stand at 75°C for 3 minutes.
	Centrifuge at 14,000 x g for 2 minutes to elute purified DNA.
8 Pure DNA	Store the DNA fragment at 4 °C or -20 °C.